



**Molecular identification of freshwater
fish species from the Commonwealth
of the Northern Mariana Islands**

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Confirming the identification of freshwater native and invasive fish species from the
Commonwealth of the Northern Mariana Islands using molecular analysis

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Prepared by:
Shelley A. James^{1,2}, Holly Bolick^{1,2} & Arnold Suzumoto^{1,3}
¹Pacific Biological Survey
²Pacific Center for Molecular Biodiversity
³Ichthyology
Bishop Museum
Honolulu HI 96817

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Introduction

It has long been recognized that DNA sequence diversity can be used to discriminate between species. Recent efforts have focused on developing a database of a single gene sequence, the mitochondrial gene cytochrome *c* oxidase subunit 1 (CO1), which is believed sufficient to differentiate the vast majority of animal species (e.g., Hebert *et al.*, 2003; Ward, *et al.*, 2005). This process is commonly referred to as barcoding. The mitochondrial cytochrome *b* (*cytb*) gene has frequently been used in molecular phylogeny studies, and can also be used for species identification (e.g., Parson *et al.*, 2000; Hsieh *et al.*, 2001; Tobe *et al.*, 2009). GenBank (<http://www.ncbi.nlm.nih.gov/Genbank/>) is the NIH genetic sequence database, consisting of more than 100 billion publicly available DNA sequences. This resource can be used to search for DNA sequences similar to that of an unknown specimen using *blast* algorithms.

Very little is known of the species that inhabit the freshwater habitats of the Commonwealth of the Northern Mariana Islands (CNMI). An extensive survey of freshwater interstitial streams on Saipan (McCagan *et al.*, 2008) found seven native species and eight species of introduced fish, but were unable to confirm the identification of many of the species due to their size and morphological similarity between species. Here we use molecular barcoding techniques to help with species identification.

Materials and Methods

Sample collection

Samples were collected from freshwater systems on Saipan, CNMI (McCagan *et al.*, 2008), frozen, and transported to the Pacific Center for Molecular Biodiversity, Bishop Museum for processing (Table 1). Upon thawing, muscle tissue was sampled and preserved in a salt-saturated solution of 20% dimethyl sulfoxide-EDTA (DMSO). Voucher specimens have been temporarily stored at -20C. Voucher DNA and tissues are housed at -80C in the Pacific Center for Molecular Biodiversity, Bishop Museum (PCMB 4840-4877).

DNA Extraction

Genomic DNA was extracted from approximately 25 mg DMSO-preserved muscle tissues using a DNeasy Blood and Tissue Kit (Qiagen Inc.) following the recommended protocol. The complete mitochondrial cytochrome *b* (*cytb*) gene (approx. 1,200 bp) of each sample was amplified via polymerase chain reaction (PCR) using the primers GLU-L 5'TGA TAT GAA AAA CCA TCG TTG-3' (Palumbi *et al.*, 1991) and H15915 5'-ACC TCC GAT CTY CGG ATT ACA AGA-3' (Aoyama *et al.*, 2001). Approximately 655bp were amplified from the 5' region of the cytochrome *c* oxidase subunit 1 (CO1) gene were amplified via PCR using the primers FishF1 5'-TCA ACC AAC CAC AAA GAC ATT GGC AC-3' and FishR1 5'-TAG ACT TCT GGG TGG CCA AAG AAT CA-3' or FishR2 5'-ACT TCA GGG TGA CCG AAg AAT CAG AA-3' for samples PCMB 4848 and 4874 (Ward *et al.*, 2005). Reactions were performed in a 50 µL solution containing 10 ng genomic DNA, 400 µM of each dNTP, 1.5 units Taq DNA polymerase (D4545, Sigma Chemical), 2 mM MgCl₂, each primer at 1 µM, and 1x PCR buffer. PCR cycling parameters were 94 C (1 min), 50 C (1 min), and 72 C (1 min) repeated for 35 cycles with a final extension of 72 C (5 min). PCR product was gel extracted

using QIAQuick Gel Extraction Kit (Qiagen Inc.). Cycle sequencing of 100 fmol double-stranded PCR product was carried out with the forward primer (GLU-L) for *cytb*, and both the forward and reverse primers for CO1 using a CEQ DTCS Quick Start Kit (Beckman-Coulter) following the recommended protocol. The products sequenced on a CEQ8000 genetic analysis system. Sequence data were subjected to nucleotide megablast (high similarity) search (<http://www.ncbi.nlm.nih.gov/BLAST/>) in order to identify the best taxon match for the resultant sequence for each specimen.

Results & Discussion:

Five of the specimens had 99-100% match for both the *cytb* and CO1 sequences within GenBank, namely PCMB 4840, 4846 and 4847 (*Chanos chanos*), PCMB 4842 (*Eleotris acanthopoma*), PCMB 4845 (*Megalops cyprinoides*), and PCMB 4848 (*Gambusia affinis*) (Table 1). PCMB 4841, preliminarily identified only as a Goby, had a 98% (*cytb*) and 99% identity with GenBank sequences for *Stiphodon elegans* (Appendix 1). *Stiphodon elegans* and *Megalops cyprinoides* have been previously reported as native to the Commonwealth of the Northern Mariana Islands (CNMI) (McCagan *et al.*, 2008). *Chanos chanos* and *Gambusia affinis* are introductions to the CNMI, and have been previously recorded from the islands. *Eleotris acanthopoma* is recorded as native to Guam, Palau, and the Western Pacific (FishBase, 2009), and would be expected to also be native in Saipan's freshwater ecosystems.

Specimens PCMB 4846 and 4847 were originally identified as a mullet species (Table 1). Molecular analysis, however, determined the two specimens to be *Chanos chanos*, and a secondary inspection of morphological characters of the voucher specimens indicated that while the specimens superficially resembled mullets, they lacked the diagnostic characteristics of the Mugilidae (A. Suzumoto, pers. comm., 2009).

Specimens PCMB 4859 and 4866 were preliminarily identified as *Poecilia reticulata* (Table 1). Both specimens had identical *cytb* and CO1 sequence data, and this confirmed the samples to be species in the genus *Poecilia*. However, no match to species level could be confirmed, most likely due to an absence of sequence data for the unknown species having been submitted to GenBank. CO1 and *cytb* sequence data were available for *Poecilia reticulata*, but the Saipan specimens had significantly different sequence data (see Appendix 2).

The three tilapia species identified in this study were *Oreochromis* species (Table 1, Appendix 3). Two of the three species had 100% match for the CO1 gene as *Oreochromis mossambicus* (PCMB 4874 and 4877). PCMB 4872, however, had 99% identity match with *Sarotherodon galilaeus* for the *cytb* gene, and 99% identity match for *Oreochromis niloticus* for the CO1 gene. This discrepancy between the two gene regions is unclear, but may be due to misidentification within the GenBank database: the CO1 gene submissions may be more reliable due to the strict identification protocols now required for barcoding projects in order to generate a standardized reference library (see Fish-Bol, <http://www.fishbol.org/>). Historically, the CNMI had aquaculture farms featuring tilapia (*O. niloticus* and *O. mossambicus*) (McCagan *et al.*, 2008). Hybrids of *O. mossambicus* x *O. niloticus* are possible, but hybridization among species can not be recognized with the molecular techniques used in this study. Mitochondrial DNA is maternally inherited, and any hybrid or subsequent generation would have the maternal species DNA only.

References:

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Table 1: Specimens analyzed in this project. Details for the collection sites can be found in McCagan *et al.* (2008). Species names in bold indicate a high probability in the maximum identity columns of genus and/or species match.

PCMB number	Collection site	Preliminary identification	cytb maximum identity	CO1 maximum identity
4840	6B	<i>Chanos chanos</i>	100% <i>Chanos chanos</i> (700bp) 84% <i>Nemacheilus putaoensis</i>	100% <i>Chanos chanos</i> (655 bp) 84% <i>Barbonymus gonionotos</i>
4841	5A	Goby	98% <i>Stiphodon elegans</i> (400 bp) 85% <i>Gillichthys</i> sp. 85% <i>Rhinogobius</i> sp.	99% <i>Stiphodon elegans</i> (626 bp) 87% <i>Sicyopterus lagocephalus</i> 86% <i>Gnatholepis</i> sp.
4842	6B	<i>Eleotris</i> cf. <i>acanthopoma</i>	100-99% <i>Eleotris acanthopoma</i> (600bp) 98% <i>Eleotris sandwicensis</i> 84% <i>Eleotris melanosoma</i> 84% <i>Eleotris fusca</i>	99% <i>Eleotris acanthopoma</i> (644 bp) 99% <i>Eleotris sandwicensis</i> 86% <i>Eleotris picta</i>
4845	6B	<i>Megalops cyprinoides</i>	99% <i>Megalops cyprinoides</i> (414 bp) 89% <i>Megalops atlanticus</i>	99% <i>Megalops cyprinoides</i> (470 bp) 82% <i>Pervagor melanocephalus</i>
4846	6A	Mullet c.f. <i>Liza vaigiensis</i>	100% <i>Chanos chanos</i> (500 bp) 84% <i>Alosa pseudoharengus</i>	100% <i>Chanos chanos</i> (642 bp) 83% <i>Barbonymus gonionotos</i>
4847	6A	Mullet	No data.	99% <i>Chanos chanos</i> (458 bp)
4848	9A	<i>Gambusia</i> sp.	100-99% <i>Gambusia affinis</i> (700bp) 92% <i>Gambusia vittata</i> 91% <i>Gambusia atrora</i>	No data.
4859	6A	<i>Poecilia reticulata</i>	97% <i>Poecilia latipinna</i> (500 bp) 94% <i>Poecilia latipunctata</i> 92% <i>Poecilia gillii</i> 85% <i>Poecilia reticulata</i>	96% <i>Poecilia petenensis</i> (649 bp) 95% <i>Poecilia mexicana</i>
4866	6B	<i>Poecilia reticulata</i>	97% <i>Poecilia latipinna</i> (500 bp) 94% <i>Poecilia latipunctata</i> 92% <i>Poecilia gillii</i> 85% <i>Poecilia reticulata</i>	94% <i>Poecilia petenensis</i> (553 bp) 93% <i>Poecilia mexicana</i>
4872	9A	Tilapia	99% <i>Sarotherodon galilaeus multifasciatus</i> (600bp) 98% <i>Sarotherodon linnellii</i>	99% <i>Oreochromis niloticus</i> (600 bp) 99% <i>Oreochromis aureus</i> 98% <i>Sarotherodon galilaeus</i>
4874	11A	Tilapia	No data	100% <i>Oreochromis mossambicus</i> (400 bp)
4877	2A	Tilapia	95% <i>Oreochromis karongae</i> (370 bp) 95% <i>Oreochromis malagarasi</i> 95% <i>Oreochromis tanganicae</i> 94% <i>Oreochromis mossambicus</i>	100-94% <i>Oreochromis mossambicus</i> (636 bp) 95% <i>Oreochromis urolepis</i> 94% <i>Oreochromis niloticus</i>

Appendix 1: Mitochondrial cytochrome *b* sequence data for *Stiphodon elegans* specimen included in this study (PCMB 4841), and data from GenBank. Dashes indicate an identical nucleotide match with *S. elegans*.

```

.....|.....|.....|.....|.....|.....|.....|.....|
      10      20      30      40      50
S. elegans AAATTGCAAA TCACGCACTA GTTGACCTCC CTGCCCTTC AAATATTTCT
4841 -----

.....|.....|.....|.....|.....|.....|.....|.....|
      60      70      80      90     100
S. elegans GCATGATGAA ACTTTGGCTC CTTTCTAGGA CTCTGCCTTG GTGCCCAAAT
4841 -----

.....|.....|.....|.....|.....|.....|.....|.....|
     110     120     130     140     150
S. elegans TGTCCTGGG CTATTTCTTG CAATACACTA TACCTCTGAC ATTGCCACAG
4841 -----

.....|.....|.....|.....|.....|.....|.....|.....|
     160     170     180     190     200
S. elegans CTTTCTCATC TGTTGCACAC ATTTGCCGTG ATGTTAACTT TGGCTGACTA
4841 -----C-----

.....|.....|.....|.....|.....|.....|.....|.....|
     210     220     230     240     250
S. elegans ATCCGGAACA TACATGCTAA TGGTGCCTCC TTCTTCTTTA TTTGCATTTA
4841 -----

.....|.....|.....|.....|.....|.....|.....|.....|
     260     270     280     290     300
S. elegans CCTGCACTTA GGGCGAGGCC TTTATTATGG CTCATACCTT TACAAAGAAA
4841 -----G-----A-----C-----

.....|.....|.....|.....|.....|.....|.....|.....|
     310     320     330     340     350
S. elegans CATGAAACAT TGGTGTAGTT CTACTACTAC TTGTAATAAT AACTGCCTTT
4841 -----

.....|.....|.....|.....|.....|.....|.....|.....|
     360     370     380     390     400
S. elegans GTAGGCTATG TTCTCCCCTG AGGACAAATA TCCTTCTGAG GGGCCACAGT
4841 -----

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Appendix 2: Mitochondrial cytochrome *b* sequence data for *Poecilia* species included in this study (PCMB 4859 & 4866), and data from GenBank for *Poecilia reticulata* (the preliminary species identification), and *P. latipinna* species (most closely matching species). Dashes indicate an identical nucleotide match with *P. latipinna*.

```

      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      10      20      30      40      50
P. reticulata -----G- -----C-- ----G-C--- ----- -A--AA-A--
P. latipinna CACCCCCTCT TAAAAATTGC AAACAATGCA CTAGTAGACC TCCCCGCCCC
4866 -----
4859 -----G-----

```

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      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      60      70      80      90     100
P. reticulata -C-A--T--T --A----- ----T----- A--T--C--- ---T-----
P. latipinna CGTCAACATC TCCGCCTGAT GAAACTTTGG CTCCCTACTA GGACTATGCC
4866 -----
4859 -----

```

```

      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      110     120     130     140     150
P. reticulata --G-T----- --T----- --T----- --T-----
P. latipinna TCATCGCCCA AATCCTAACC GGCTTATTTT TAGCAATACA CTACACCTCT
4866 -A-----
4859 -A-----

```

```

      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      160     170     180     190     200
P. reticulata --T--T--T- ----- --C--G-- -----A--
P. latipinna GACATCTCCA TAGCATTCTC ATCTGTAACC CACATTTGTC GAGATGTTAA
4866 -----T----- -----G-- --T-----
4859 -----T----- -----G-- --T-----

```

```

      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      210     220     230     240     250
P. reticulata T--T----- --T--T---- -C--A--C-- T--T--A--- -----G--T-
P. latipinna CTACGGATGA CTCATCCGCA ATATGCATGC CAACGGGGCC TCCCTATTCT
4866 -----
4859 -----T--A--- -----

```

```

      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      260     270     280     290     300
P. reticulata -C--T--C- ----C-G-- ----- -AT----- T--G-----
P. latipinna TTATCTGCAT CTACTIONACAT ATTGGACGAG GCCTATACTA CGGTTCCCTAC
4866 -----
4859 -----

```

```

      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      310     320     330     340     350
P. reticulata C-G-T----- -G----- -----T--A --C--AT-T- -T--A-----
P. latipinna TTATACAAAG AAACATGAAA CATTGGCGTT ATTCTCCTAC TACTTGTTAT
4866 -----
4859 -----

```

```

      .....|.....| .....|.....| .....|.....| .....|.....| .....|.....|
      360     370     380     390     400
P. reticulata ----- -C----- -T--A-----

```


Molecular identification of freshwater fish species from CNMI

```

P. latipinna AATAACTGCT TTTGTAGGAT ACGTTCTCCC ATGAGGACAA ATATCCTTCT
4866          -----
4859          -----
                |         |         |         |         |         |
                410       420       430       440       450
P. reticulata ---T--A--  ---T---  --T-----A-  -----C---A-----
P. latipinna GAGGCGCCAC TGTAATCACC AACCTCCTCT CTGCTGTACC TTACGTAGGC
4866          -----
4859          -----
                |         |         |         |         |         |
                460       470       480       490       500
P. reticulata --TG-C----  -----  -----A-  -----A--
P. latipinna GACTCTCTG TCCAATGAAT CTGAGGGGGA TTTTCAGTTG ACAACGCTAC
4866          -----
4859          -----

```

Appendix 3: Mitochondrial cytochrome *b* sequence data for an unknown *Tilapia* species included in this study (PCMB 4877), and data from GenBank for the most closely matching species of *Oreochromis*. Dashes indicate an identical nucleotide match with unknown specimen. CO1 data indicated a 100% match with *Oreochromis mossambicus*.

	5	15	25	35	45
4877	CTAAAAATTG	CAAACGACGC	ACTAGTTGAC	CTCCCAGCCC	CCTCAAACAT
<i>O. karongae</i>	-----	-----	-----	-----	-----
<i>O. malagarasi</i>	-----	-----	-----	-----	-----
<i>O. tanganicae</i>	-----	-----	-----	-----	-----
<i>O. mossambicus</i>	-----	-----	-----	-----	-----

	55	65	75	85	95
4877	TTCCGTTTGA	TRAAACTTTG	GATCTCTACT	AGGCCTCTGT	CTAGCCGCCC
<i>O. karongae</i>	-----	-g-----	-g-----	-----	-----
<i>O. malagarasi</i>	c-----	-g-----	-----	---g-----	-----
<i>O. tanganicae</i>	-----	-g-----	-----	-----	-----
<i>O. mossambicus</i>	-----	-g-----	-----	-----	-----a--

	105	115	125	135	145
4877	AAATCCTGAC	AGGCCTTTTC	CTAGCCATAC	ACTATACTTC	CGACATCGCC
<i>O. karongae</i>	-----	-----	-----	---c-----	-----
<i>O. malagarasi</i>	-----	-----	-----	-----c--	-----
<i>O. tanganicae</i>	-----	-----t	-----	---c--c--	-----
<i>O. mossambicus</i>	-----	-----t	-----	---c--c--	-----

	155	165	175	185	195
4877	ACAGCCTTCT	CCTCCGTTGC	TCATATTTGT	CRAGATGTAA	ACTACGGCTG
<i>O. karongae</i>	-----	-----c--	c--c-----	-g-----	-----
<i>O. malagarasi</i>	-----	-----c--	c--c-----	-g--c-----	-----
<i>O. tanganicae</i>	-----	-----c--	c--c-----	-g-----	-----t-----
<i>O. mossambicus</i>	-----	-----c--	c--c-----	-g--c-----	-----

	205	215	225	235	245
4877	ACTCATTCTA	AACATACATG	CCAACGGCGC	ATCTTTCTTC	TTCATTTGTA
<i>O. karongae</i>	-----g-	-----	-----	-----	-----
<i>O. malagarasi</i>	-----g-	-----	-----	-----	-----
<i>O. tanganicae</i>	-----g-	-----	-----	-----	-----c-
<i>O. mossambicus</i>	-----g-	-----	-----	-----	-----c-

	255	265	275	285	295
4877	TTTATCTCCA	CATCGGACGA	GGCTTATATT	ACGGCTCTTA	CCTGTACAAA
<i>O. karongae</i>	-----	t--t-----	---c-----	-----	-----
<i>O. malagarasi</i>	-----	---t-----	---c-----	-----	-----
<i>O. tanganicae</i>	-----	---t-----	---c-----	-----	-----
<i>O. mossambicus</i>	-----	---t-----	---c-----	-----	-----

	305	315	325	335	345
4877	GAAACCTGAA	ACATTGGAGT	CATTCTCCTC	CTCCTAACTA	TAATAACAGC
<i>O. karongae</i>	-----	-----	t-----	-----	-----

Molecular identification of freshwater fish species from CNMI

```

O. malagarasi ----- t-----c-----
O. tanganicae ----- t--c-----g-----
O. mossambicus ----- catcctcctc -----

      ....|....| ....|....|
      355      365
4877  CTTTCGTAGGC TACGTCCTCC
O. karongae -----t-----
O. malagarasi -----t-----
O. tanganicae -----tc-a-----
O. mossambicus ---t-----t-----

```